Research Article

Biosynthesis and Characterization of TiO₂ Nanoparticles by Lactococcus lactis ssp. lactis

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ArticleInfo ABSTRACT

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Lactococcus lactis ssp. *lactis* isolated from raw milk was used for titanium dioxide (TiO₂) nanoparticles biosynthesis. Biosynthesized TiO₂ nanoparticles were characterized using UV.Vis spectroscopy, Atomic Force Microscopy (AFM) (1.97 nm), X-ray diffraction (XRD) apparatus, Field Emission Scanning Electron Microscopy (FE-SEM), Energy dispersive X-ray analysis (EDX) spectra and Fourier Transform Infrared Spectroscopy (FTIR). Result was 408.21 cm⁻¹ that belong to anatase Titania. *L. lactis* ssp. *lactis* isolates had the ability to synthesize TiO₂ nanoparticles, the characterization results presented that the biosynthesized nanoparticles were at wavelength (344-347) nm; approving the formation of anatase phase of TiO₂ NPs; spherical crystals, with particles average diameter of 47.22 nm.

KEYWORDS: Probiotics bacteria, Lactococcus spp., Biosynthesis, TiO₂ nanoparticles.

الخلاصة

اختبرت قابلية بكتيريا المعززات الحيوية العائدة للجنس Lactococcus lactis ssp. lacti المعزولة من الحليب الخام لإنتاج الجسيمات النانوية لثنائي اوكسيد التيتانيوم (TiO2). وصفت هذه الجسيمات النانوية المصنعة باستخدام جهاز مطياف الأشعة المرئية وفوق البنفسجية (UV.Vis)، وجهاز حيود الأشعة السينية (XRD)، مجهر القوة الذرية (AFM) (T.97) نانوميتر، جهاز التحليل الطيفي بالأشعة تحت الحمراء (FTIR) حيث كانت احد القمم الناتجة 20.10 سم⁻¹ والتي تعود لطور الاناتيز الجسيمات التيتانيوم النانوية، المجهر الإلكتروني الماسح بالانبعاثات الالكترونية (FE-SEM) ومطيافية المشتنة المشت الطاقة (EDX). L. lactis ssp. lactis يحد الطول الموجي (Ater) نائريوم النانوية المعنية وصيف جسيمات الناتيز وكسيد التيتانيوم النانوية، المجسيمات المصنعة كانت عند الطول الموجي (Ater) ومطيافية الأشعة السينية المشتنة موحد للبلورات النانوية نوع معمله وبمعدل حجم (47.22) نانوميتر.

INTRODUCTION

Probiotics are defined as living microorganisms that might be found in cultured milk and fermented food; they can be used in infant's food preparations [1]. Control on and reduce the serum cholesterol are some of the benefits of probiotic bacteria besides enhancing food products nutritional values, suppressing antibiotic-associated diarrhea and preventing gut infections [2, 3].

Lactococcus lactis subsp. *lactis* was isolated from milk or milk fermented products and plant material too. It was used for the fermentation of milk and buttermilk as well as wide types of cheeses [4]. *L. lactis* enhanced the microbial balance of the gastrointestinal tract by suppressing the growth of hydrogen sulfide (H₂S) producing microorganisms in the intestine of mice [1]. *L. lactis* was used in vaccines as a live vector. It is considered as a promising candidate for heterologous expression of protein and biotechnological application [5].

Green synthesis of biogenic nanoparticles (NPs) from plant or microbial sources became an emergent field due to being eco-friendly, fast, simple, safer, energy-efficient, cheap, and less toxic [6]. Bacterial applications for biomanufacturing of NPs have several advantages such as the short incubation time and easy control, for example, bacteria are considered as an excellent tool for extracellular synthesis for silver and gold nanoparticles [7, 8].

Many studies indicated that probiotic bacteria produce a lot of metal nanoparticles/oxide nanoparticles involving Gd₂O₃, Sb₂O₃, Ag, CdS, Se, Au, ZnO, and TiO₂ [9]. Ibrahem *et al.* (2014)





used a biological method for the synthesis of TiO_2 NPs besides studying the effect of different culture media on TiO_2 nanoparticles that were biosynthesized by *Lactobacillus crispatus* [10].

Nanoparticles reveal distinctive, catalytic, optical, magnetic, and electronic properties that are not the same as bulk metals [11]. The importance of TiO₂ NPs was related to their chemical stability; photocatalytic activity; low-priced; non-toxicity besides being Generally Recognized as Safe (GRAS) [12].

Studies on metal oxide NPs proved that they act as potential antimicrobial agents [13]. Depending on the foodstuff, the type of packaging matrix is of importance; the addition great of metal nanoparticles as (Ag, Cu, Hg, and TiO₂) to polymer thermal matrices. mechanical properties, resistance, gas barrier properties, and antimicrobial properties is considered to be improved in comparison with the usual polymers [14]. This study aimed to use green synthesis of TiO₂ NPs by probiotic's bacteria Lactococcus lactis ssp. lactis.

MATERIALS AND METHODOLOGY

Lactococcus lactis ssp. lactis isolates

Buffalo and cows' raw milk was used as a source for *Lactococcus lactis* ssp. *lactis* isolation, their identification was dependent on the cultural, microscopical, and biochemical tests [15]; in addition to the Vitek 2 system.

Biosynthesis of TiO2 Nanoparticles

Pure culture of L, lactis ssp. lactis (9×10^8 CFU/ml), inoculated at 2% in De Man Rogosa and Sharpe (MRS) broth flasks; then were incubated at 30 °C, for 24 hrs. under anaerobic conditions. TiO2 NPs detected by color-changing (light to dark brown) and sediment production [16]. The product was later to centrifuge transferred tubes; the centrifugation process was done at 5000 rpm for 5 min. After that, the pellet was washed with deionized water; this step was repeated several times until pure solution was acquired. After the final centrifugation, the pellet was dried for 1 hr. at 50 °C until TiO₂ nanoparticles were converted to powder formula [17].

Characterization of Biosynthesized TiO₂ nanoparticles

The characterization of biosynthesized TiO_2 nanoparticles was done by using: UV.Vis

spectroscopy [18]; Atomic Force Microscopy (AFM) [19]; X-ray diffraction (XRD) [20]; Fourier Transform Infrared Spectroscopy (FTIR) [21]; Field Emission Scanning Electron Microscopy (FE-SEM) and Energy dispersive X-ray analysis (EDX) [21].

RESULTS AND DISCUSSION

Six isolates of L. lactis ssp. lactis were identified from raw milk samples. L. lactis ssp. lactis isolates had the ability for synthesizing TiO₂ NPs and this result was observed by the sediment formation and color-changing from light brown to dark. The detected absorption spectrum corresponds with those gained with TiO₂ nanoparticles synthesized by Lactococcus isolates, with an absorption band in the UV range and its cut-off wavelength was (344-347) nm (Figure 1); approving the formation of anatase phase of TiO₂ NPs [16]. The absorption band was at 406 nm for TiO2 nanoparticles biosynthesized extracellularly by Lactobacillus johnsonii in the anatase phase [22]. The UV spectra indicated a distinct absorption peak revealing anatase phase similar to the peak detected of TiO₂ NPs synthesized via Staphylococcus aureus at 324 nm wavelength [19].



igure 1. The UV.VIs spectroscopy for *Lactococcus lactis* ssp. *lactis*.

The topographical 2D and 3D images with AFM microscope for TiO_2 nanoparticles biosynthesized by *L. lactis* ssp. *lactis* result was (1.97 nm) as in Figure 2. Atomic Force Microscopy depicted anatase formation formulas of TiO_2 nanoparticles as well surface morphology as a result of incidence of some aggregates and single particles. Roughness with no linear trend; which verified that the highest TiO_2 concentrations would lead to smoother layers of materialization [19].



Figure 2. AFM of TiO₂ nanoparticles biosynthesized by Lactococcus lactis ssp. lactis.

The anatase phase is known as the most active photocatalytic one [23]. Furthermore, layers of anatase phase typically work as a super hydrophilic surface [24]. Attributable to the greater photocatalytic activity, anatase has the highest number of industrial applications as the most commercially used form [25].



Figure 3. XRD pattern of TiO₂ nanoparticles biosynthesized by *Lactococcus lactis* ssp. *lactis*.

Titanium dioxide nanoparticles surface micrograph biosynthesized by L. lactis ssp. lactis was made up of a uniform distribution of spherical crystals in nanostructure; the size was 47.22 nm. The FE-SEM result was shown in (Figure 4) for biosynthesized TiO₂ NPs; this image evidently gives the size and physical morphology of particles. The surface morphology (shape and size) of NPs was analyzed by SEM varies in size of particles and might be correlated to the formation of TiO₂ nanoparticles at different times [21]. SEM result, spherical shapes with diverse particles diameter of 70-90 nm for the synthesized TiO₂ NPs, these differences may be associated with the aggregation phenomena of particles [26]. Decreasing particles size is inversely proportional to surface volume; therefore, the smaller particle size quickly penetrates the bacterial surface, which leads to the process of decomposition [27].



Figure 4. FESEM image of TiO₂ nanoparticles biosynthesized by *Lactococcus lactis* ssp. *lactis*.

L. lactis ssp. *lactis* EDX scales for biosynthesized TiO_2 NPs powders confirmed the existence of (Ti, O). No other peaks for other elements were detected in the spectrum, which confirmed the results of biosynthesized nanoparticles purity (Figure 5). This result verified that the particles were crystalline and metallic nanoparticles of TiO_2 [28].



Figure 5. EDX of biosynthesized TiO₂ nanoparticles by *Lactococcus lactis* ssp. *lactis*.





The absorption peaks shown in the FTIR technique were within the spectrum associated to TiO₂ nanoparticles. The main IR topographies of TiO₂ NPs biosynthesized by L. lactis ssp. lactis showed at 408.21 cm⁻¹ and belong to anatase Titania (Figure 6); the range of anatase Titania wavenumber (400- 1000) cm⁻¹ as Ti-O-Ti bridge [29, 30] and 1060.88 cm⁻¹ corresponds to the Ti-OH as mentioned in [31] the 1068 cm⁻¹ band was attributed to Ti-OH bond. Another peak was measured at (1646.01, 3273.41) cm⁻¹, the first peak was attributed to the stretching of titanium carboxylate; while the second one was for O-H stretching mode of surface and the adsorbed water molecules; other vibrational bands were observed at 2929.47 cm⁻¹ for L. lactis ssp. lactis, and assigned to C-H asymmetric stretching vibrations [30]. Xing et al., 2021 noticed a similar band around 2920 cm⁻¹ corresponding to the C-H stretching vibration of alkyl and aldehyde groups for TiO₂ nanoparticles [31].

Laisney *et al.*, 2021 mentioned that; TiO_2 NPs could be found at 1630 cm⁻¹ corresponding to the Ti (TiO₂)-O (H₂O), vibrations between the titanium or oxygen from the TiO₂ surface and the oxygen from the hydroxyl layer [32]. The increased bridging hydroxyls on the surface reduce the positive charges of TiO₂ and cause particle aggregation, negatively affecting catalytic performance [33].



Figure 6. FTIR results of biosynthesized TiO₂ nanoparticles by *Lactococcus lactis* ssp. *Lactis*.

CONCLUSIONS

Iraqi probiotics isolates belong to *Lactococcus lactis* ssp. *Lactis* had the ability to synthesize pure anatase TiO_2 nanoparticles with particles diameter of 47.22 nm.

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REFERENCES

- [1] F. Zendeboodi, N. Khorshidian, A. M. _Mortazavian and A. G. Cruz. _ Probiotic: conceptualization from a new approach. Current Opinion in Food Science.32: 103-123, 2020.
- [2] M. G. Gareau , P. M. Sherman and W. A. Walker. Probiotics and the gut microbiota in intestinal health and disease. Nature reviews. Gastroenterology and hepatology. 7(9), 503-514, 2010.
- [3] M. Kechagia, D. Basoulis, S. Konstantopoulou, D. Dimitriadi, K. Gyftopoulou, N. Skarmoutsou and E. M. Fakiri. Health benefits of probiotics: a review. ISRN nutrition. 481651: 1-7, 2013.
- [4] R. W. Hutkins. Microbiology and Technology of Fermented Foods. Blackwell Publishing Professional, Ames, IA, pp. 3–14, 2006.
- [5] M. Azizpour, S. D. Hosseini, P. Jafari and N. Akbary. Lactococcus lactis: A new strategy for vaccination. Avicenna journal of medical biotechnology. 9(4): 163–168, 2017.
- [6] P. Velusamy, G. V. Kumar, V. Jeyanthi, J. Das and R. Pachaiappan. Bio- inspired green nanoparticles: synthesis, mechanism, and antibacterial application. Toxicological research. 32(2): 95–102, 2016.
- [7] M. Gomathy and K. Sabarinathan. Microbial mechanisms of heavy metal tolerance-a review. Agricultural Reviews. 31(2): 133-138, 2010.
- [8] A. Ayesha. Bacterial synthesis and applications of nanoparticles. Nano Science and Nano Technology an Indian Journal. 11(2):119-149, 2017.
- [9] K. K. Darani, A. G. Cruz, M. R. Mozafari, Z. Abdi and N. Ahmadi. Biosynthesis of metal nanoparticles by probiotic bacteria. Letters in Applied NanoBioScience. 8(3): 619-626, 2019.
- [10] K. H. Ibrahem, J. A. S. Salman and F. A. Ali. Effect of titanium nanoparticles biosynthesis by *Lactobacillus crispatus* on urease, hemolysin & biofilm forming by some bacteria causing recurrent UTI in Iraqi women. European Scientific Journal, ESJ. 10(9): 324-338, 2014.
- [11] H. J. Prabu and I. Johnson. Plant-mediated biosynthesis and characterization of silver nanoparticles by leaf extracts of *Tragia involucrata*, *Cymbopogon citronella*, *Solanum verbascifolium* and *Tylophora ovata*. Karbala International Journal of Modern Science. 1(4), 237-246, 2015.
- S. Liu, D. Tao, H. Bai and X. Liu. Cellulose-nanowhiskertemplated synthesis of titanium dioxide/cellulose nanomaterials with promising photocatalytic abilities.
 J. Appl. Polymer Polysaccharides. 25(1): 282–290, 2012.
- [13] R. K. Singh, J. C. Knowles and H. W. Kim. Advances in nanoparticle development for improved therapeutics

delivery: nanoscale topographical aspect. Journal of tissue engineering. 10: 1-9, 2019.

- [14] S. Paidari and S. A. Ibrahim. Potential application of gold nanoparticles in food packaging: a mini review. Gold Bulletin. 54: 31–36, 2021.
- [15] F. J. Carr, D. Chill and N. Maida. The lactic acid bacteria: a literature survey. Critical Rev. Microbiol. 28(4): 281-370, 2002.
- [16] K. E. Alzahrani, A. A. Niazy, A.M. Alswieleh, R. Wahab, A. M. El-Toni and H.S. Alghamdi. Antibacterial activity of trimetal (CuZnFe) oxide nanoparticles. International journal of nanomedicine. 13: 77–87, 2017.
- [17] M. Mishra, J. S. Paliwal, S. K. Singh, E. Selvarajan, C. Subathradevi and V. Mohanasrinivasan. Studies on the inhibitory activity of biologically synthesized and characterized zinc oxide nanoparticles using *Lactobacillus sporogens* against *Staphylococcus aureus*. Journal of Pure and Applied Microbiology. 7(2): 1263-1268, 2013.
- [18] P. Anandgaonker, G. Kulkarni, S. Gaikwad and A. Rajbhoj. Synthesis of TiO₂ nanoparticles by electrochemical method and their antibacterial application. Arabian Journal of Chemistry. 12(8):1815-1822, 2015.
- [19] K. S Landage, G. K Arbade, P. Khanna and C. J. Bhongale. Biological approach to synthesize TiO₂ nanoparticles using *Staphylococcus aureus* for antibacterial and antibiofilm applications. Journal of Microbiology & Experimentation. 8(1):36–43. 2020.
- [20] T. Theivasanthi and M. Alagar. Titanium dioxide (TiO₂) nanoparticles - XRD analyses – An insight. Chemical Physics. 1: 1-10, 2013.
- [21] A. Jha and K. Prasad. Biosynthesis of metal and oxide nanoparticles using *Lactobacilli* from yoghurt and probiotic spore tablets. Microbial Biotechnology Journal. 5(3): 285-291, 2010.
- [22] H. A. Al-Zahrani, A. A. El-Waseif and D. E. El-Ghwas. Biosynthesis and evaluation of TiO₂ and ZnO nanoparticles from in vitro stimulation of *Lactobacillus johnsonii*. Journal of Innovations in Pharmaceutical and Biological Sciences (JIPBS). 5(1): 16-20, 2018.
- [23] M. R. Mohammadizadeh, M. Bagheri, S. Aghabagheri and Y. Abdi. Photocatalytic activity of TiO₂ thin films by hydrogen DC plasma. Applied Surface Science. 350: 43-49, 2015.
- [24] E. G. Mariquit, W. Kurniawan, M. Miyauchiro and H. Hinode. Effect of addition of surfactant to the surface hydrophilicity and photocatalytic activity of immobilized Nano-TiO₂ thin films. Journal of Chemical Engineering of Japan. 48(10): 856-861, 2015.
- [25] A. Khalid, P. Ahmad, A. Alharthi, S. Muhammad, M. U. Khandaker, M. R. I. Faruque, I. U. Din and M. A.

Alotaibi. Unmodified titanium dioxide nanoparticles as a potential contrast agent in photon emission computed tomography. Crystals. 11(2): 171- 181, 2021.

- [26] G. Gohari, A. Mohammadi and A. Akbari. Titanium dioxide nanoparticles (TiO₂ NPs) promote growth and ameliorate salinity stress effects on essential oil profile and biochemical attributes of *Dracocephalum moldavica*. Scientific reports. 10(1): 912- 926, 2020.
- [27] M. Aravind, M. Amalanathan and M. S. M. Mary. Synthesis of TiO₂ nanoparticles by chemical and green synthesis methods and their multifaceted properties. SN Applied Sciences. 3(409): 1-10, 2021.
- [28] T. Santhoshkumar, A. A. Rahuman, C Jayaseelan, G. Rajakumar, S. Marimuthu, A. V. Kirthi, K. Velayutham, J. Thomas, J. Venkatesan and S.K. Kim. Green synthesis of titanium dioxide nanoparticles using *Psidium guajava* extract and its antibacterial and antioxidant properties. Asian Pacific Journal of Tropical Medicine. 7(12):968-976, 2014.
- [29] P. Praveen., G. Viruthagiri and N._ Shanmugam. Structural, optical and morphological analyses of pristine titanium di-oxide nanoparticles – Synthesized via sol-gel route. Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy. 117: 622-629, 2014.
- [30] D. Dodoo-Arhin, F. P. Buabeng, J. M. Mwabora, P. N. Amaniampong, H. Agbe, E. Nyankson, D. O. Obada and N.Y. Asiedu. The effect of titanium dioxide synthesis technique and its photocatalytic degradation of organic dye pollutants. Heliyon. 4(7): 1- 23, 2018.
- [31] Y. Xing, X. Li, X. Guo, W. Li, J. Chen, Q. Liu, Q. Xu, Q. Wang, H. Yang, Y. Shui and X. Bi. Effects of different TiO₂ nanoparticles concentrations on the physical and antibacterial activities of chitosan-based coating film. Nanomaterials (Basel, Switzerland). 10(7): 1365-1384, 2020.
- [32] J. Laisney, A. Rosset, V. Bartolomei, D. Predoi, D. Truffier-Boutry, S. Artous, V. Bergé, G. Brochardd and I. Michaud-Soret. TiO₂ nanoparticles coated with bioinspired ligands for the safer-by-design development of photocatalytic paints. Environmental Science: Nano. 8(1): 297-310, 2021.
- [33] C. Y. Wu, K. J. Tu, J. P. Deng, Y. S. Lo and C. H. Wu. Markedly enhanced surface hydroxyl groups of TiO₂ nanoparticles with superior water-Dispersibility for photocatalysis. Materials (Basel, Switzerland). 10(5): 566-581, 2017.



